



Genekor Medical S.A. | 52, Spaton Ave., 15344, Gerakas, Athens, Greece ,G.E.MI. nr: 0007856001000
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 Scientific Director: George Nasioulas PhD

SAMPLE INFORMATION

Name :	-	Date Received :	-
Medical ID :	-	Date of Report :	-
Date of Birth :	-	Req. Physician :	-
Location :	-	Barcode :	-
Material :	WHOLE PERIPHERAL BLOOD	Reason of referral:	Referral for hypotonia, long bone fractures

Whole Exome analysis (WES) by Next Generation Sequencing

Results associated with the reason of referral

PATHOGENIC VARIANT IDENTIFIED

Gene	Variant	Clinical Significance	Zygosity
TRIP4	NM_016213.5:c.832C>T, p.(Arg278*)	Pathogenic variant	Homozygous



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- George Nasioulas, PhD Molecular Biologist, Scientific Director, AMKA:26025301255

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Variants Details

TRIP4, Exon 7, NM_016213.5:c.832C>T, p.(Arg278*)

ClinGen

HPO

ClinVar

This is a single base substitution, replacing Arginine with a Termination codon in the TRIP4 gene (p.Arg278*). This results in the production of a truncated, non-functional protein. This variant has been described in international bibliography in homozygosity or compound heterozygosity in 2 families with affected individuals presented with prenatal-onset spinal muscular atrophy (SMA), multiple congenital contractures (arthrogryposis multiplex congenita), respiratory distress, and congenital bone fractures. ([PMID: 26924529](#)). This particular variant has been described mutation database ClinVar ([Variation ID: 224632](#)). For these reasons this variant has been classified as pathogenic. According to international guidelines it is recommended that relatives of the patient are tested for the above mutation.

The *TRIP4* gene encodes a subunit of the tetrameric ASC-1 transcriptional cointegrator complex. The other subunits include ASCC1 (614215), ASCC2 (614216), and ASCC3 (614217). The complex associates with transcription factors or with nuclear receptors and can bidirectionally affect the link between receptor and transcription machinery, either as corepressor or coactivator. This complex also likely participates in pre-mRNA processing and regulation of splicing. Pathogenic/likely pathogenic variants are cause spinal muscular atrophy with congenital bone fractures 1 and are inherited in the recessive manner.



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Methodology

Genomic DNA was extracted from the sample under investigation and was analysed by a solution based capture approach using the target enrichment panel KAPA HyperExome Probes, 43Mb, Roche. Sequencing was carried out using MGI technology. Reads were aligned to the reference sequence (GRCh37), and sequence changes were identified and interpreted in the context of a single clinically relevant transcript. Unless otherwise stated, this assay targets all coding regions of the indicated transcripts and 10 base pairs of flanking intronic sequences. All targeted regions were sequenced with $\geq 10x$ depth.

Based on the available patient information, the following diagnostic algorithm was used.

- The genes described in the OMIM and HPO databases were selected as genes associated with the patient phenotype.
- Variant classification was performed according to the ACMG AND AMP guidelines (PMID: 25741868)
- Analysis of the mutations described in the HGMD, the mutations with damaging effect (frameshift, nonsense, missense, splicing mutations etc.) as well as de novo mutations was obtained.
- All clinically significant observations were confirmed by Sanger Sequencing.

*Note:

Every molecular test has an internal 0,5-1% chance of failure. This is due to rare molecular events and factors related to the preparation and analysis of the samples. Unless otherwise stated, this assay targets all coding regions of the indicated transcripts and 10 base pairs of flanking intronic sequences. Therefore, this method cannot detect variants in deep intronic or enhancer/promoter regions.

The method used achieves 99% sensitivity and specificity for single nucleotide variants and insertions and deletions.



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Details about non-pathogenic variants

All individuals carry DNA changes (i.e., variants), and most variants do not increase an individual's risk of the disease. When identified, variants of uncertain significance (VUS) are reported. Findings of variants of uncertain significance (VUS) in autosomal recessive diseases are not reported unless they co-exist with another finding (pathogenic, likely pathogenic or VUS in the same gene). Benign variants (Polymorphisms) are not reported and available data indicate that these variants most likely do not cause increased risk of the disease. Present evidence does not suggest that non-clinically significant variant findings be used to modify patient medical management beyond what is indicated by the personal and family history and any other clinically significant findings.

Genes Analyzed (Table 1)

ACAN	ACP5	ACTB	ACTG1	ACVR1	ADAMTS10	ADAMTS17	ADAMTS2	ADAMTSL2
AFF4	AGA	AGPS	AIFM1	AKT1	ALPL	ALX1	ALX3	ALX4
AMER1	AMMECR1	ANKH	ANKRD11	ANO5	ANTXR2	ARCN1	ARHGAP31	ARID1A
ARID1B	ARSB	ASAH1	ASCC1	ASXL1	ATP6V0A2	ATP7A	ATR	B3GALT6
B3GAT3	B4GALT7	BCOR	BCS1L	BGN	BHLHA9	BICD2	BLM	BMP1
BMP2	BMPER	BMPR1B	BRAF	BRCA2	BRIP1	BSCL2	C2CD3	CA2
CANT1	CASR	CBL	CC2D2A	CCDC47	CCDC8	CDC42	CDC45	CDC6
CDH11	CDH3	CDKN1C	CDT1	CENPE	CENPJ	CEP120	CEP152	CEP290
CEP63	CHCHD10	CHST14	CHST3	CHSY1	CKAP2L	CLCN5	CLCN7	COG4
COL10A1	COL11A1	COL11A2	COL1A1	COL1A2	COL27A1	COL2A1	COL3A1	COL5A1
COL5A2	COL9A1	COL9A2	COL9A3	COMP	CREB3L1	CREBBP	CRIP1	CRLF1
CRTAP	CSF1R	CSPP1	CTSK	CUL7	CWC27	CYP27B1	CYP2R1	DCTN1
DDR2	DDRGK1	DHCR24	DHCR7	DHODH	DLL3	DLL4	DLX3	DLX5
DMP1	DNAJB2	DNAJC21	DNMT3A	DOCK6	DSE	DVL1	DVL3	DYM
DYNC1H1	DYNC2H1	DYNC2LI1	EBP	EDN1	EDNRA	EFL1	EFNB1	EFTUD2
EIF2AK3	EIF4A3	ENAM	ENPP1	EOGT	EP300	ERCC4	ERF	ESCO2
EVC	EVC2	EXOSC2	EXOSC3	EXOSC8	EXT1	EXT2	EXTL3	EZH2
FAM111A	FAM20A	FAM20C	FAM83H	FANCA	FANCB	FANCC	FANCD2	FANCE
FANCF	FANCG	FANCI	FANCL	FANCM	FBN1	FBN2	FBXO38	FERMT3
FGD1	FGF10	FGF23	FGF9	FGFR1	FGFR2	FGFR3	FIG4	FKBP10
FKBP14	FLNA	FLNB	FN1	FTO	FUCA1	FZD2	GALNS	GALNT3
GCM2	GDF3	GDF5	GDF6	GH1	GHR	GHRHR	GHSR	GJA1

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GLB1	GLI1	GLI2	GLI3	GMNN	GNAI3	GNAS	GNPAT	GNPTAB
GNPTG	GNS	GORAB	GPC6	GSC	GUSB	GZF1	HAAO	HDAC4
HDAC8	HES7	HESX1	HEXA	HOXA11	HOXA13	HOXD13	HPGD	HRAS
HSPA9	HSPB1	HSPB3	HSPB8	HSPG2	IARS2	IDH2	IDS	IDUA
IFIH1	IFITM5	IFT122	IFT140	IFT172	IFT43	IFT52	IFT80	IFT81
IGF1	IGF2	IGFALS	IGHMBP2	IHH	IL1RN	INPPL1	INSR	INTU
IRS1	KAT6B	KCNJ2	KIAA0586	KIAA0753	KIF22	KIF7	KL	KMT2A
KRAS	KYNU	LARP7	LAS1L	LBR	LEMD3	LFNG	LHX3	LHX4
LIFR	LMNA	LMX1B	LONP1	LPIN2	LRP4	LRP5	LTBP2	LTBP3
LZTR1	MAFB	MAN2B1	MANBA	MAP2K1	MAP2K2	MAP3K7	MATN3	MBTPS2
MECOM	MEGF8	MEOX1	MESP2	MET	MGP	MKS1	MMP13	MMP2
MMP9	MNX1	MSX2	MYCN	MYH3	MYO18B	NAGLU	NANS	NBAS
NEK1	NEU1	NF1	NFIX	NIPBL	NKX3-2	NOG	NOTCH1	NOTCH2
NPR2	NRAS	NSD1	NSDHL	OBSL1	OCRL	OFD1	ORC1	ORC4
ORC6	OSGEP	OSTM1	OTX2	P3H1	P4HB	PALB2	PAM16	PAPSS2
PAX3	PCGF2	PCNT	PCYT1A	PDE3A	PDE4D	PEX14	PEX19	PEX5
PEX7	PGM3	PHEX	PIGV	PIK3CA	PISD	PITX1	PITX2	PLCB4
PLEKHG5	PLEKHM1	PLK4	PLOD1	PLOD2	PLS3	POC1A	POLA1	POLR1A
POLR1C	POLR1D	POLR3A	POLR3B	POP1	POR	POU1F1	PPIB	PPP3CA
PRKAR1A	PRMT7	PROP1	PTDSS1	PTH1R	PTHLH	PTPN11	PUF60	PYCR1
RAB23	RAB33B	RAD21	RAD51C	RAF1	RALA	RASA2	RBBP8	RBM8A
RBPJ	RECQL4	REEP1	RIPPLY2	RIT1	RMRP	RNU4ATAC	ROR2	RPGRIP1L
RRAS	RSPRY1	RTTN	RUNX2	SALL1	SALL4	SBDS	SC5D	SCO2
SEC24D	SERPINF1	SERPINH1	SETBP1	SETD2	SF3B4	SFRP4	SGMS2	SGSH
SH3BP2	SH3PXD2B	SHH	SHOC2	SHOX	SKI	SLC10A7	SLC17A5	SLC26A2
SLC29A3	SLC34A3	SLC35D1	SLC39A13	SLC5A7	SLCO2A1	SLX4	SMAD2	SMAD3
SMAD4	SMARCA2	SMARCA4	SMARCAL1	SMARCB1	SMARCE1	SMC1A	SMC3	SMN1
SMN2	SNRPB	SNX10	SOS1	SOST	SOX11	SOX2	SOX3	SOX9
SP7	SPARC	SRCAP	SRP54	STAG2	STAMPB	STAT5B	SUMF1	TAB2
TALDO1	TAPT1	TBCE	TBX15	TBX19	TBX2	TBX3	TBX4	TBX5
TBX6	TBXAS1	TCF12	TCIRG1	TCOF1	TCTN3	TGDS	TGFB1	TGFB3
TGFBR1	TGFBR2	THPO	TMEM165	TMEM216	TMEM38B	TMEM67	TNFRSF11A	TNFRSF11B
TNFSF11	TONSL	TOP3A	TP63	TRAF3IP1	TRAPPC2	TREM2	TRIM37	TRIP11
TRIP4	TRMT10A	TRPS1	TRPV4	TRPV6	TTC21B	TUBGCP6	TWIST1	TYROBP
UBA1	UFSP2	VAPB	VDR	VIPAS39	VRK1	WDR19	WDR35	WNT1

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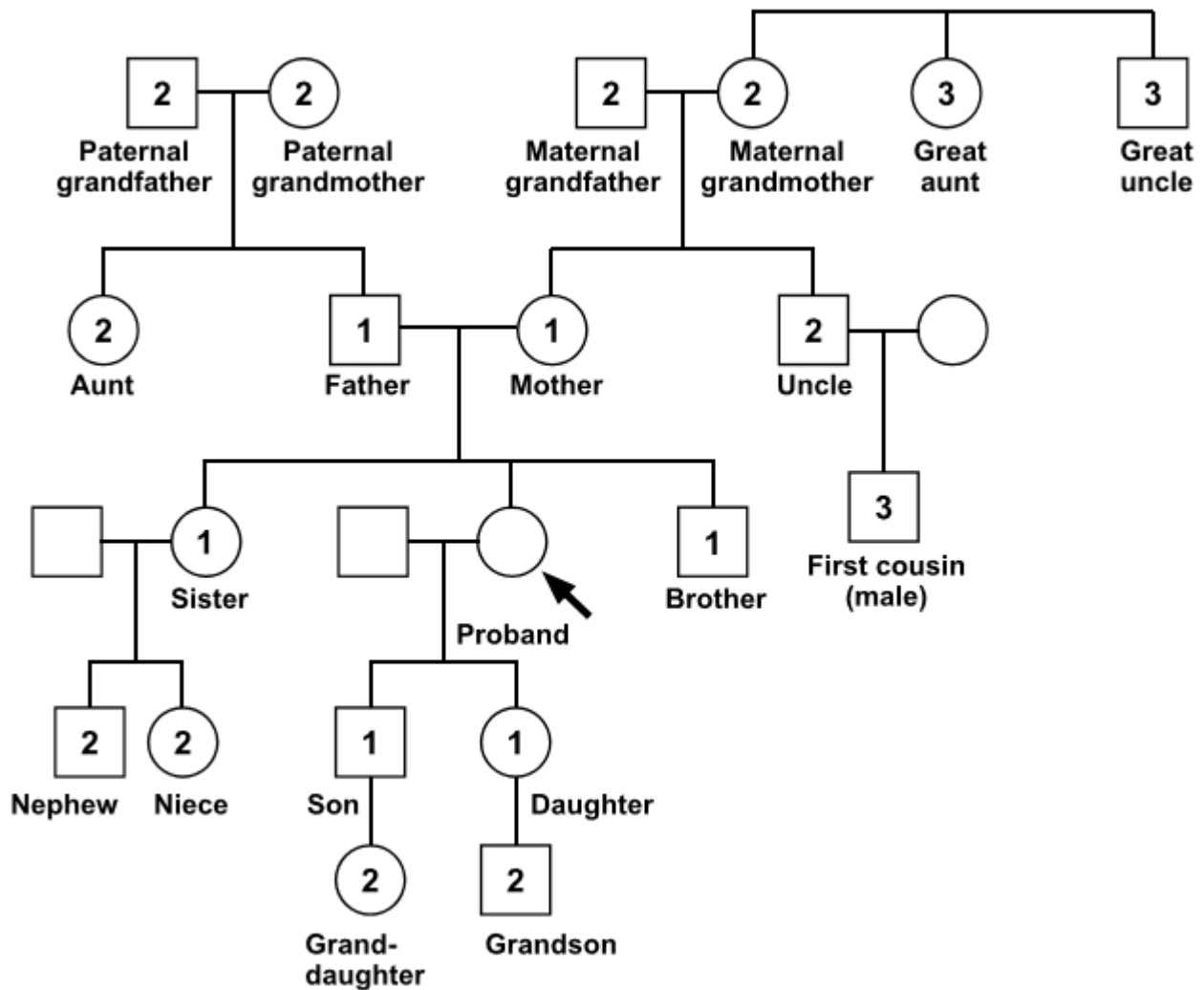
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WNT10B	WNT5A	WNT7A	XRCC2	XRCC4	XYLT1	XYLT2	ZMPSTE24	ZSWIM6

Family tree



Note: The information shown on the family tree has been provided by the patient and not by medical records.



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Literature

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