



# 1709P - Cascade genetic testing utilized only in 31% of initial families with pathogenic variants in breast cancer genes

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# Genekor

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## INTRODUCTION

Hereditary cancer predisposition syndromes are responsible for approximately 5-10% of all diagnosed cancer cases. Identification of genetic predisposition using germline testing, is usually followed by targeted variant testing in family members, ensuring a cost-effective approach for identifying high-risk individuals. Consequently, this has significant implications for treatment decisions, risk-reducing interventions, and cancer screening and prevention.

## OBJECTIVES

The aim of this study is to examine the clinical use and implementation of cascade family testing (CFT) in the families of breast cancer patients with pathogenic/likely pathogenic variants (PVs/LPVs).

## MATERIALS AND METHODS

We performed a retrospective analysis of 1785 individuals referred to our laboratory for genetic testing using a multigene panel. A capture-based method NGS technology was used for the analysis of 43 genes involved in hereditary cancer predisposition. Sequencing was carried out using the Illumina NGS technology. Reads were aligned to the reference sequence (GRCh37), and sequence changes were identified and interpreted in the context of a single clinically relevant transcript. The capture-based approach allowed for computational analysis of CNVs from NGS data. Cascade testing in 117 individuals was performed with Sanger sequencing or MLPA. The schematic representation of the workflow can be seen below:

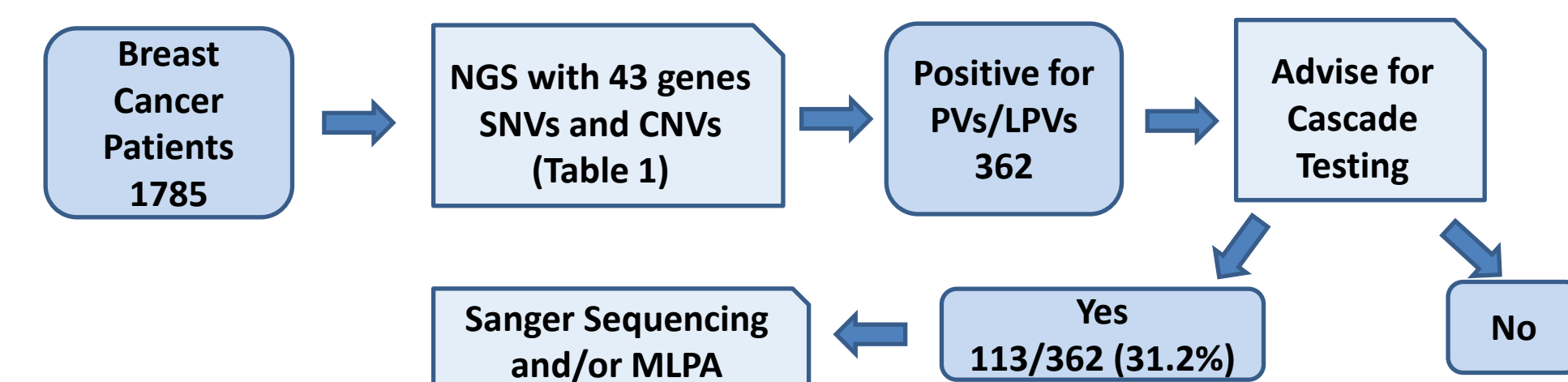


Figure 1. Schematic representation of the workflow used in this study.

Table 1. Genes analyzed.

| Gene                      | Reference sequence   | Gene                   | Reference sequence |
|---------------------------|----------------------|------------------------|--------------------|
| APC                       | NM_000038            | MSH6 <sup>*2</sup>     | NM_000179          |
| ATM <sup>*1</sup>         | NM_000051            | MUTYH <sup>*2</sup>    | NM_001128425       |
| AXIN2                     | NM_004655            | NBN <sup>*1</sup>      | NM_002485          |
| BARD1 <sup>*1</sup>       | NM_000465            | NF1                    | NM_000267          |
| BMPRI1A                   | NM_004329            | NTHL1                  | NM_002528          |
| BRCA1 <sup>*1,2</sup>     | NM_007294            | PALB2 <sup>*1,2</sup>  | NM_024675          |
| BRCA2 <sup>*1,2</sup>     | NM_000059            | PMS2                   | NM_000535          |
| BRIP1 <sup>*1</sup>       | NM_032043            | POLD1 (Exons 8-13)     | NM_001256849       |
| CDH1                      | NM_004360            | POLE (Exons 1-14)      | NM_006231          |
| CDK4                      | NM_000075            | PPP2R2A <sup>*1</sup>  | NM_002717          |
| CDKN2A (p14ARF, p16INK4a) | NM_000077, NM_058195 | PTEN                   | NM_000314          |
| CHEK2 <sup>*1,2</sup>     | NM_007194            | RAD50 <sup>*1,2</sup>  | NM_005732          |
| EPCAM <sup>*2</sup>       | NM_002354            | RAD51B <sup>*1</sup>   | NM_133509          |
| FANCA <sup>*1</sup>       | NM_000135            | RAD51C <sup>*1,2</sup> | NM_058216          |
| FANCL <sup>*1</sup>       | NM_001114636         | RAD51D <sup>*1,2</sup> | NM_002878          |
| FANCM <sup>*1</sup>       | NM_020937            | RET                    | NM_020975          |
| HOXB13:c.251G>A p.(G84E)  | NM_006361            | SMAD4                  | NM_005359          |
| MEN1                      | NM_000244            | SMARCA4                | NM_001128849       |
| MLH1 <sup>*2</sup>        | NM_000249            | STK11                  | NM_000455          |
| MRE11 <sup>*1</sup>       | NM_005591            | TP53 <sup>*2</sup>     | NM_000546          |
| MSH2 <sup>*2</sup>        | NM_000251            | VHL                    | NM_000551          |
| MSH3                      | NM_002439            |                        |                    |

<sup>\*1</sup> Genes of the homologous recombination (HR) complex

<sup>\*2</sup> Unless otherwise noted analysis of large rearrangement was performed on the following genes:

BRCA1, BRCA2, CHEK2, EPCAM (Exons 8, 9), MLH1, MSH2, MSH6, MUTYH, PALB2, RAD50 (Exons 1, 2, 4, 10, 14, 21, 23 and 25), RAD51C, RAD51D, and TP53.

## RESULTS

In a cohort of 1785 patients with breast cancer, PVs/LPVs were found in 362 patients (20.3%). In specific, 52.2%, 25.1%, and 22.7% of positive patients had findings in high-, moderate- and low- penetrance cancer susceptibility genes, respectively. Although, CFT was advised in all families, 117 individuals from 113 families (31.2%) continued with genetic testing. The mean ages of probands and first-degree relatives tested were 46 and 40 years (p<0.0001), respectively. Of the first-degree relatives who underwent CFT, 70% were female and the 105/117 (89.7%) were asymptomatic (Table 2). The median time to cascade testing was 9 months.

Among the 117 tested individuals, 42.7%, 36.7% and 20.6% were offspring, siblings, and parents of probands, respectively (Fig.2). In total, the familial PV/LPV was detected in 53.0% (62/117) of first-degree relatives tested. Our data suggest that CFT was mostly undertaken (104/113, 91.4%) in families with positive findings in high-risk genes, although these represent only 55.0% (104/189) of initial families with PV/LPV in high-risk genes (Fig 3).

Table 2. Information of individuals who performed CFT.

|                     | N=117       |
|---------------------|-------------|
| Female, n (%)       | 82 (70%)    |
| Male, n (%)         | 35 (30%)    |
| Mean Age, years     | 40          |
| Asymptomatic, n (%) | 105 (89.7%) |
| Symptomatic, n (%)  | 12 (10.3%)  |

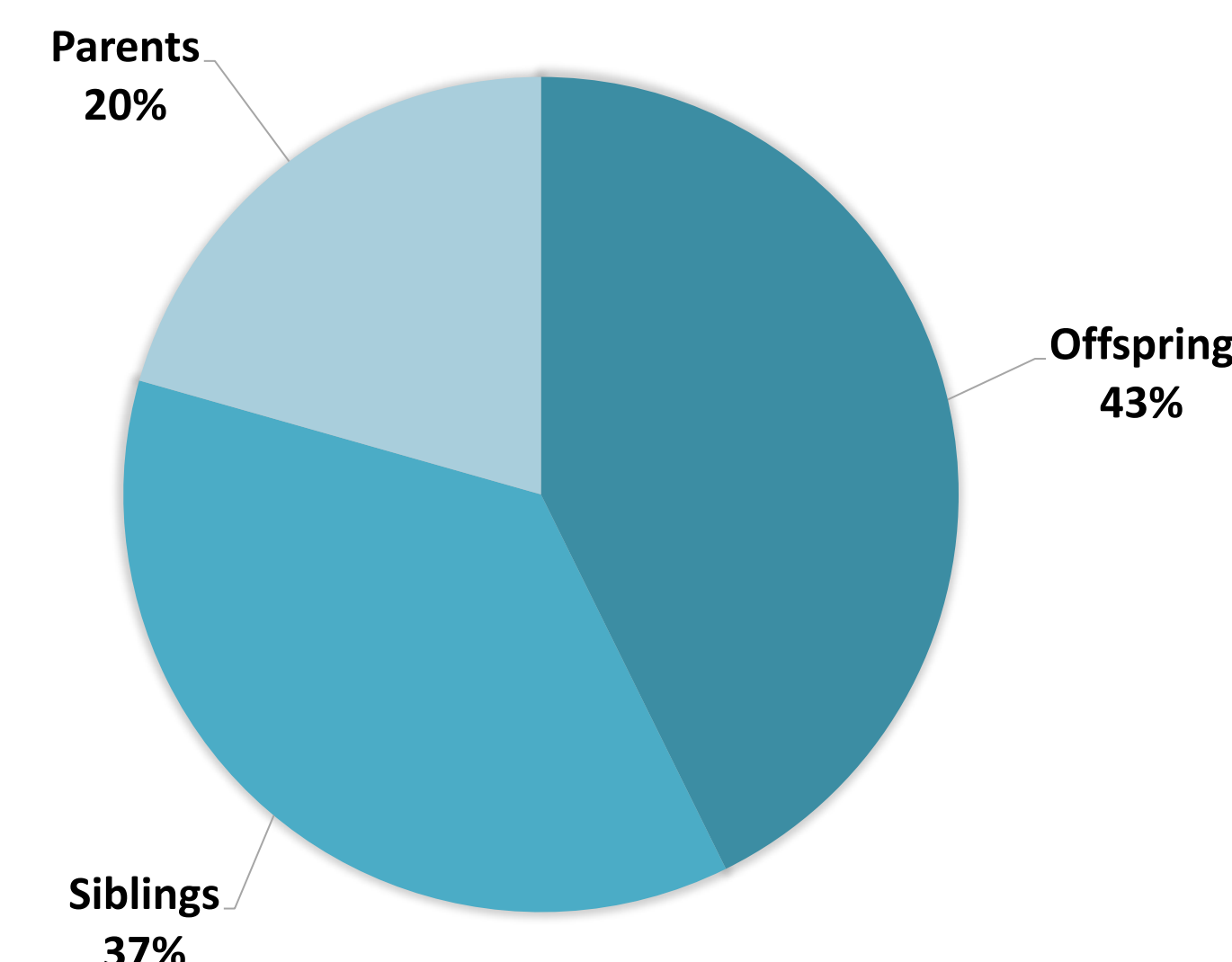


Figure 2. Percentage of first-degree relative categories undergoing cascade testing.

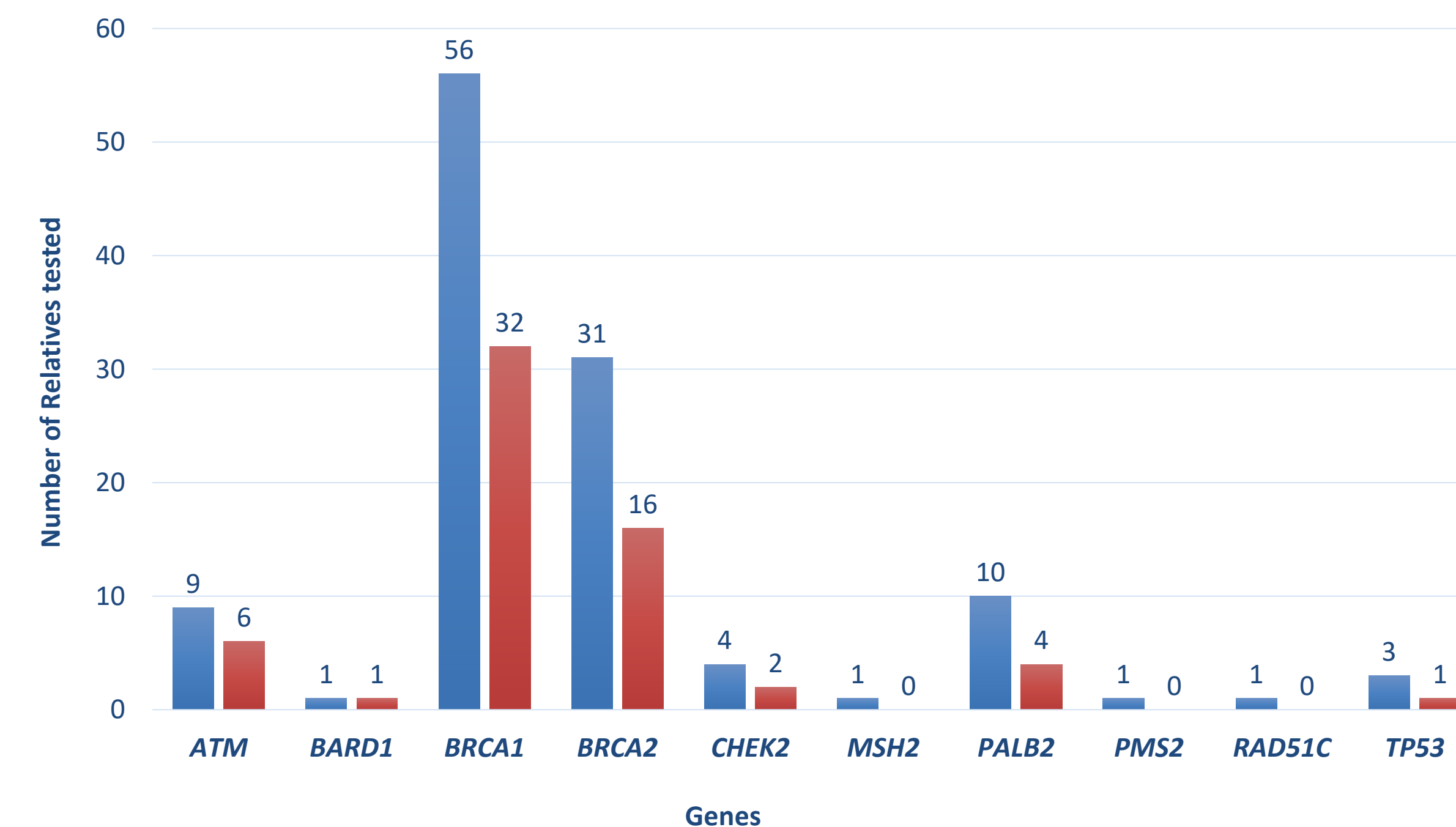


Figure 3. Number of relatives who pursued cascade testing with a negative (blue color) or positive (red color) results.

## DISCUSSION

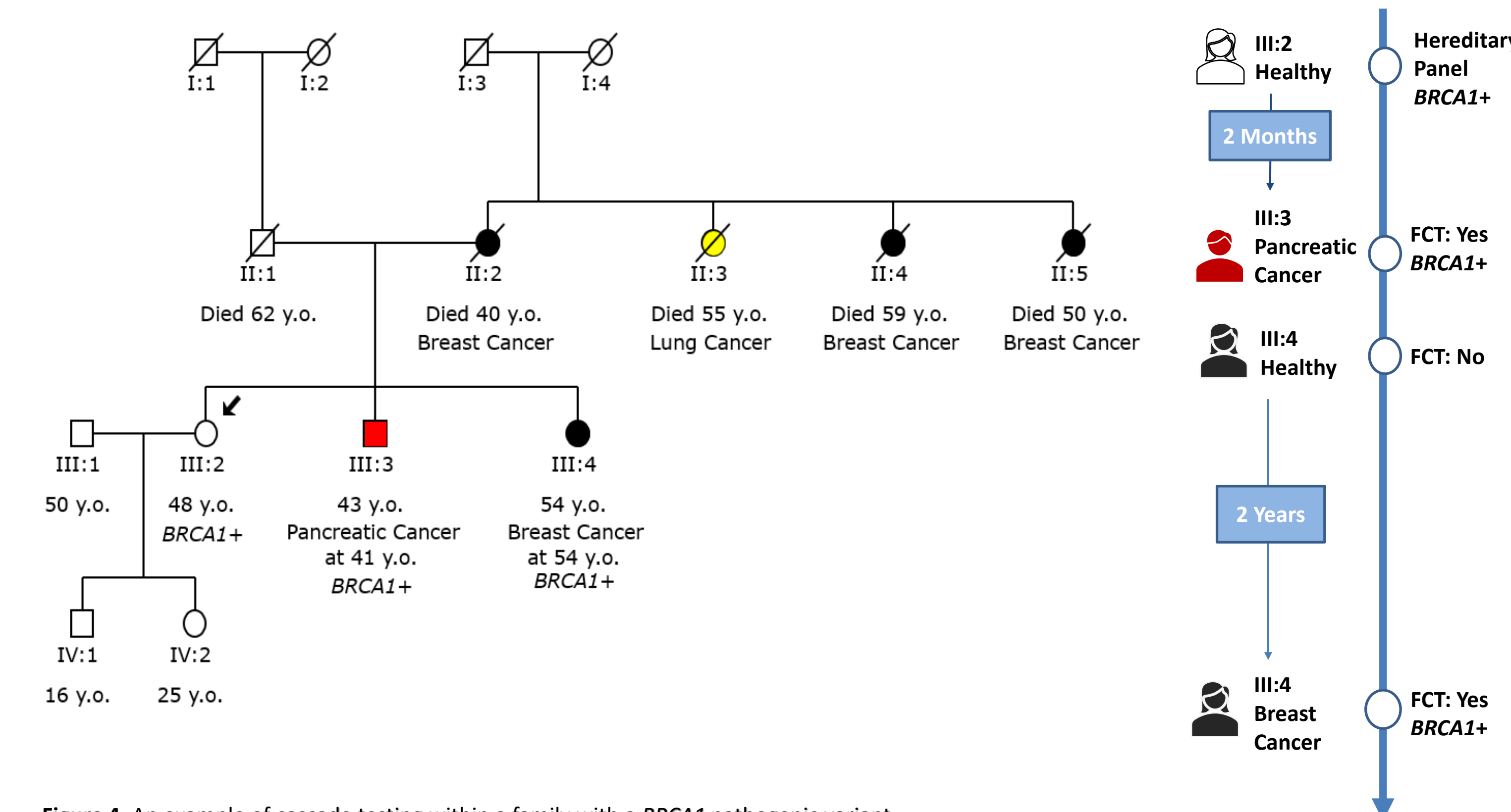


Figure 4. An example of cascade testing within a family with a BRCA1 pathogenic variant.

In this family the initial proband (III:2) was tested positive for a pathogenic variant in the BRCA1 gene. The reason of referral was the strong family history of breast cancer. After the announcement of the results the brother of the proband (III:3) who developed pancreatic cancer was tested positive for the specific variant in the BRCA1 gene. The sister of the proband (III:4) who was healthy at that time did not consent to cascade family testing. Two years later, she (III:4) was diagnosed with breast cancer and she decided to be tested for the specific variant in the BRCA1 gene. The analysis revealed that she carried the pathogenic variant in the BRCA1 gene.

## CONCLUSIONS

- Our results underline the fact that CFT is underutilized, even in families with PVs/LPVs in high-risk genes.
- Cascade testing can be a powerful tool for primary cancer prevention towards the identification of at-risk family members.
- Additional efforts should be targeted to the implementation of genetic counseling approaches leading to better family education and communication of genetic testing results.

## REFERENCES

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